Optimising Foliar Inputs

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Why Foliar Feed?

- Foliar fertilisation strategies can achieve:
 - -Higher nutrient use efficiency (& economics)
 - -Improve yield, quality and metabolism of crops
 - -Reduce the negative impact on the environment
 - -Potentially enhance consumer health benefits



agriculture

Niu et al (2021). doi.org/10.1007/s42729-020-00346-

* Sarkar et al (2021). doi.org/10.3390/agriculture11040372

Nutrient	Efficiency (%)
Nitrogen	30-50
Phosphorus	15-20
Potassium	50-60
Sulphur	8-12
Zinc	2–5
Iron	1–2
Copper	1–2
Manganese	1–2
Boron	2–3
Molybdenum	2–5



¹ Niu et al (2021). doi.org/10.1007,

Benefits to Soil?

- Worldwide, 20% of cultivated land and 33% of irrigated land is salt-affected and degraded.
- Foliars have been shown to:
 - Reduce soil nitrates
 - Reduce soil salinity
 - Reduce soil acidification



LAND

Problems with Uptake?

- Soil nutrient supply limited by: –Abiotic stresses: temperature, soil moisture, salinity, pH extremes.
 - -Biotic stresses: pest and disease, soil biological activity.



Foliars Enhance Soil Uptake?

- Foliar NH₄⁺ increased soil N uptake (cotton)
- Foliar K promoted soil K uptake (potato)
- Foliar urea enhanced soil N and P uptake (potato)
- Mechanism?
 - Foliar nutrients can be translocated to roots and increase root biomass and hence soil uptake.

* Niu et al (2021). doi.org/10.1007/s42729-020-00346-3

– Enhanced root exudation.



Cons

- Variable/inconsistent responses have been seen.
- Upper limit to units/ha that can be applied.
- Poor translocation/mobility.
- Shorter lived benefits and follow up applications likely required.
- Possibility of burning if solution too concentrated.
- Weather restrictions wind, rainfall, temperature, humidity.





Uptake Pathways





Huurna/ or Explorental Boo

Strategies for probing absorption and translocation of foliarapplied nutrients





* Eichert & Fernández (2011). doi.org/10.1016/B978-0-12-384905-2.00004-2

Strategies for probing absorption and translocation of foliarapplied nutrients



ANNALS OF BOTANY

* Li et al (2019). doi.org/10.1093/aob/mcy13





* Kopittke et al (2020). doi.org/10.1:



Nondestructive diagnostics of magnesium deficiency based on distribution features of chlorophyll concentrations map on cucumber leaf









the plant journal

dez et al (2021), doi.org/10.1111/tr

Foliar water and solute absorption: an update





C-based inputs, chelates, adjuvants



Chelation

- Chelated minerals generally have:
 - Neutral charge
 - Better absorption (by root or foliar)
 - Better translocation within the plant
 - Lower salt index/less burning
 - Slower release rate (varies)
- Amino chelates are particularly useful.
 Fish etc











* McCov et al (2020), doi.org/10.3390/agronomv10030358

* Stiegler et al (2011). doi.org/10.2135/cropsci2010.06.0377



Humic Substances



Characteristic	Humic Acids	Fulvic Acids
Molecular Weight	10,000-100,000 Daltons	1000-10,000 Daltons
0	% of organic	components
Carbon	50-60	40-50
Hydrogen	4-6	4-6
Nitrogen	2-6	1-3
Oxygen	30-35	44-50
Sulfur	0-2	0-2



Plant Response: Application

- Nozzle
- Droplet size
- Pressure
- Droplet deflection
- Runoff
- Drift
- Surface area coverage
- Forward speed

Plant Response: Crop

- Crop stage
- Canopy Structure

* Sible et al (2021). doi.org/10.3390/agronomy110712

- Leaf Area Index
- Leaf Chemistry
- Leaf Shape
- Cuticle
- Surfaces waxes
- Leaf hairs, spines etc
- Abiotic and biotic stresses





Plant Response: Environment

- Humidity
 - Time of day (stomata & pores open)
 - 70%+ humidity is ideal
 - Rapid drying can lead to re-crystallization
- Temperature
- Max 28 C (~25 C)
- Wind
- 3-15 km/h
- Drought
 - Early vs Late plant stress

Plant, Cell & Environment

Unravelling foliar water uptake pathways: The contribution of stomata and the cuticle

PCE

Abstract Plants can absorb water through their leaf surfaces, a phenomenon commonly referred to as foliar water uptake (FWU). Despite the physiological importance of FWU, the pathways and mechanisms underlying the process are not well known. Using a novel experimental approach, we parsed out the contribution of the stomata and the cuticle to FWU in two species with Mediterranean (Prunus duicis) and temperate (Pruva communis) origin. The hydraulic parameters of FWU were derived by analysing mass and water potential changes of leaves placed in a fog chamber. Leaves were previously treated with abscisic acid to force stomata to remain closed, with fusioccin to remain open, and with water (control). Leaves with open stomata rehydrated two times faster than leaves with closed stomata and attained approximately three times higher maximum fluxes and hydraulic conductance. Based on FWU rates, we propose that rehydration through stomata occurs primarily via diffusion of water vapour rather than in liquid form even when leaf surfaces are covered with a water film. We discuss the potential mechanisms of FWU and the significance of both stomatal and cuticular pathways for plant productivity and survival.

zmán-Delgado *et al* (2021), doi.org/10.1111/pce.14041

the plant journal





* https://grdc.com.au/__data/assets/pdf_file/0023/142583/grdc_fs_spray-practical-tips_low-res-pdf.pdf.pdf



"...it is crucial to develop new methods to increase nitrogen use efficiency (NUE); and it is estimated that, even a 1% increase in NUE could save \$1.1 billion US dollar per annum"

* Kant et al (2011). doi.org/10.1093/jxb/erq297 * Stuart et al (2014). doi.org/10.1016/j.landusepol.2013.08.011

Mechanisms of Efficiency Gains?

- Improved and even coverage vs sporadic granules
- Less nutrient antagonisms/competition vs soil applied
- Less ammonia volatilisation, less nitrate leaching
- Maintain nutrient supply during sub-optimal soil conditions
- Metabolic shortcutting of efficient N forms more energy left for leaf and root growth





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 - Foliar nutrients can be translocated to roots and increase root biomass and hence soil uptake.
 - Enhanced root exudation.

Foliar Efficiencies

- Michigan State University
 - 1950's
 - Tukey and Wittwer

* Autio & Bramlage, https://ag.umass.edu/sites/ag.umass.edu/files/fact-sheets/pdf/folcalcium.pdf

- Radioactive P demonstrated rapid uptake
- 95% was taken up as compared to 10-20% soil applied
- 8-10 or 20 times more efficient

* Niu et al (2021). doi.org/10.1007/s42729-020-00346-3

*Tukey et al (1952) doi.org/10.1126/science.116.3007.167 *Bukovac & Wittwer (1957) doi.org/10.1104/pp.32.5.428

Foliar Efficiencies

Nutrient	Crop type	Foliar ratio	Soil ratio	Source
Zinc (ZnSO4)	Annual crops	1	12	Lingle & Holmberg (1956)
Phosphorus (H3PO4)	beans, tomatoes	1	20	Wittwer, et al. (1957)
Iron (FeSO4)	Grain, sorghum	1	25	Withee & Carlson (1959)
Magnesium (MgSO4)	Grain, sorghum	1	100	Krantz (1962)
Magnesium (MgSO4)	Celery	1	50-100	Johnson, et al. (1957, 1961)
Magnesium (MgSO4) Magnesium (MgSO4)	Grain, sorghum Celery	1 1	100 50-100	Krantz (1962) Johnson, et al. (1957,

Approximate ratios of amounts required for comparable crop response.

* https://advancednutrients.com.au/wp-content/uploads/2021/05/Foliar-Fertilisers-White-Paper.pdf

Foliar Calcium Sprays for Apples

Increase in fruit calcium obtained from different treatments. The numbers in parentheses are the actual pounds of calcium applied per acre, assuming that trees required 300 gal dilutes pray per acre. All toliar treatments were madewith CaC2, "The3spray, 5-spray, and 2-spray treatments applied a yearly total of 74, 40, and 18 pounds of technicalgrade CaC2, per acre. Gypsum was applied at approximately oneton per acre, annually.

Treatment	Increasein fruit calcium concentration (ppm)
8 foliar sprays (22 lbs calcium/acre)	45
5 foliar sprays (12 lbs calcium/acre)	25
2 foliar sprays (5 lbs calcium/acre)	10
Gypsum on soil (400 lbs calcium/acr	e) 12



Foliar versus soil phosphorus (P) application for improving P use efficiency in wheat and maize in calcareous soils

0.68

90 135 LSD_{in}

SP × FP

ABTRACT PARTNET PRE-NAME PRE-NAME

* Rafiullaha et al (2021). doi.org/10.1080/01904167.202

Table 3. Total uptake and P use efficiency of maize and wheat crop as affected by different levels of phosphorus both soil and foliar applied P. Total P uptake (mg pot⁻¹) P use efficiency (%) Treatments Maize Wheat Maize Wheat Soil applied P (mg P kg soil) 122.62 c 45.06 b 4.43 2.31 160.08 b 179.86 a 13.34 67.59 a 75.94 a 8.65 7.90 4.16 20 LSD_{10.055} Foliar P (mM KH₂PO₄) 0 45 118.35 c 148.18 b 163.68 ab 186.52 a 15.40 45.84 c 59.44 b 64.52 ab 81.66 a 9.98

0.86

37.96 19.88 14.21

19.96 10.50 7.92

888

Urease inhibitor reduces N losses and improves plantbioavailability of urea applied in fine particle and granular forms under field conditions

> A field lysimeter/mini plot experiment was established in a silt loam soil near Lincoln, New Zealand, to investigate the effectiveness of urea fertilizer in fine particle application (FPA), with or without the urease inhibitor N-(n-butyl) thiophosphoric triamide (NBPT - "Agrotain"), in decreasing nitrogen (N) losses and improving N uptake efficiency. The five treatments were: control (no N) and "N-labelled urea, with or without NBPT, applied to lysimeters or mini plots (unlabelled urea), either in granular form to the soil surface or in FPA form (through a spray) at a rate equivalent to 100 kg N ha-. Gaseous emissions of ammonia (NH₃) and nitrous oxide (N₂O), nitrate (NO₃-) leaching, herbage dry-matter (DM) production, N-response efficiency, total N uptake and total recovery of applied \circ N in the plant and soil varied with urea application method and with addition of NBPT. Urea with NBPT, applied in granular or FPA form, was more effective than in application without NBPT: N₂O emissions were reduced by 7–12%, NH₂ emissions by 65–69% and NO₃-leaching losses by 36–55% compared with granular urea. Urea alone and with NBPT, applied in FPA form increased herbage DM production by 27% and 38%, respectively. The N response efficiency increased from 10 kg DM kg $^\circ$ of applied N with granular urea to 19 kg DM kg $^\circ$ with FPA urea and to 23 kg DM kg $^\circ$ with FPA urea plus NBPT. Urea applied in FPA form resulted in significantly (P < 0.05) higher «N recovery in the shoots compared with granular treatments and this was improved further when urea in FPA form was applied with NBPT. These results suggest that applying urea with NBPT in FPA form has potential as a management tool in mitigating N losses, improving N-response efficiency and increasing herbage DM production in intensive grassland systems.



agronomy

Comparing Soil vs. Foliar Nitrogen Supply of the Whole Fertilizer Dose in Common Wheat

Abstract: Late-season N application through foliar spraying is recognized as an efficient agronomic practice for improving grain quality in common wheat, although the major part of N is still supplied by soil fertilization. This study assessed the impact of various N doses entirely applied by repeated foliar sprayings on wheat growth, yield and quality, in comparison with conventional soil fertilization management with a recommended dose of 160 kg N ha⁻¹ as ammonium nitrate (C-M). Doses of 96. 104 and 120 kg N ha-1 as both UAN (urea-ammonium-nitrate) and urea applied by foliar spraying were evaluated in a 2-year field trial in Northern Italy in a silty loam soil with 1.7% organic matter. Here, it was demonstrated that the canopy greenness was similar in all treatments, with slight grain yield increases by the lowest foliar N dose vs. C-M. The higher N foliar doses mainly improved the grain protein content and both high- and low-molecular-weight glutenin subunits (HMW-GS, LMW-GS), particularly with urea. It is concluded that in our fertile soil, managing N fertilization exclusively through foliar spraying is feasible without compromising grain yield and ameliorating quality at the same time. Improved nutrient use efficiency and beneficial environmental effects are also expected by reducing the nitrogen load on the agricultural fields by 25-40%.

* Ferrari et al (2021). doi.org/10.3390/agronomy11112138

Year	Fertilizer	Treatment Yield DW (kg ha ⁻¹)		3 ha-1)	Harvest Index (%)		TSW (g)		Testing Weight (kg hL ⁻¹)	
		0N	5570 ± 9.70		34.3 ± 0.51		28.9 ± 0.26		80.9 ± 0.73	
		C-M	6407 ± 76.5	b	35.7 ± 1.11	a	29.0 ± 0.69	а	80.5 ± 0.23	b
	TIAN	F-96	6635 ± 107.1	ab (+4)	35.9 ± 0.83	a (+1)	28.8 ± 0.43	a (-1)	81.7 ± 0.38	ab (+1
	UAN	F-104	6789 ± 76.2	a (+6)	36.8 ± 1.36	a (+3)	30.3 ± 1.26	a (+5)	82.4 ± 0.49	a (+2)
2018-2019		F-120	6357 ± 121.2	b (-1)	33.7 ± 3.24	a (-6)	27.8 ± 2.39	a (-9)	80.6 ± 0.90	ab (=)
		C-M	6386 ± 41.7	ab	33.1 ± 0.96	a	27.7 ± 0.72	а	79.5 ± 0.19	b
	LIDEA	F-96	6527 ± 96.5	a (+2)	35.8 ± 0.69	a (+8)	29.4 ± 0.36	a (+6)	81.9 ± 0.36	a (+3)
	UKEA	F-104	6193 ± 95.6	b (-3)	36.0 ± 1.47	a (+9)	29.1 ± 0.44	a (-1)	80.4 ± 0.69	b (+1)
		F-120	6524 ± 69.0	a (+2)	33.9 ± 0.62	a (+2)	28.9 ± 0.66	a (-1)	81.8 ± 0.31	a (+3)
		0N	5914 ± 758.4		41.0 ± 1.69		35.5 ± 0.57		80.9 ± 0.56	
		C-M	6129 ± 436.0	a	39.9 ± 1.75	a	34.5 ± 0.51	a	82.5 ± 0.19	b
2019-2020	LIDEA	F-96	6828 ± 286.8	a (+11)	44.7 ± 0.36	a (+12)	38.3 ± 0.64	a (+10)	83.2 ± 0.38	ab (+1
	OKEA	F-104	6214 ± 643.9	a (+1)	39.9 ± 3.72	a (=)	35.1 ± 2.77	a (+2)	82.8 ± 0.38	ab (=)
		F-120	6259 ± 287.7	a (+2)	41.3 ± 3.14	a (+3)	35.2 ± 1.51	a (+2)	83.8 ± 0.38	a (+1)

		Treatment									
Year	Date	Phenological	UAN				UF	EA			
		Stage	0N	C-M	F-96	F-104	F-120	C-M	F-96	F-104	F-1
	20 October 2018	Pre-sowing	32 (s)	32 (
	25 February 2019	Tillering (ZDS 26)		58 (s)	16 (f)	8 (f)	8 (f)	58 (s)	16 (f)	8 (f)	8 (
2018-2019	21 March 2019	Stem elongation (ZDS 37)		58 (s)	16 (f)	16 (f)	16 (f)	58 (s)	16 (f)	16 (f)	16
	24 April 2019	Booting (ZDS 40)	-	-	16 (f)	32 (f)	32 (f)	-	16 (f)	32 (f)	32
	7 May 2019	Flowering (ZDS 62)	-	12 (f)	16 (f)	16 (f)	32 (f)	12 (f)	16 (f)	16 (f)	32
	22 October 2019	Pre-sowing	32 (s)	-	-	-	-	32 (s)	32 (s)	32 (s)	32 (
	25 February 2020	Tillering (ZDS 27)	-	-	-	-	-	58 (s)	16 (f)	8 (f)	8 (
2019–2020	28 March 2020	Stem elongation (ZDS 37)	-	-				58 (s)	16 (f)	16 (f)	16
	24 April 2020	Booting (ZDS 40)	-	-					16 (f)	32 (f)	32
	7 May 2020	Flowering (ZDS 62)	-	-	-	-	-	12 (f)	16 (f)	16 (f)	32
	Total N dose		32	160	96	104	120	160	96	104	12
N sa	ving (%) vs. C-M tre	atment	80%	Ref.	40%	35%	25%	Ref.	40%	35%	25

* Ferrari et al (2021). doi.org/10.3390/agronomy11112138

* Ferrari et al (2021). doi.org/10.3390/agronomy11112138

Table 1. Dates and growth stages of N application (kg ha^{-1}).

* Ferrari et al (2021)	doi.org/10.3390	/agronomv11112138

Year	Fertilizer	Treatment	GPC	(%)	Zeleny II	ndex (%)	Wet Glu	ten (%)
		0N	11.4 ± 0.53		33.5 ± 2.12		27.6 ± 1.31	
		C-M	14.0 ± 0.64	а	42.8 ± 5.88	а	31.4 ± 1.68	ab
	UAN	F-96	13.1 ± 0.37	a (-6)	35.9 ± 2.40	a (-16)	29.4 ± 0.80	b (-6)
2010 2010		F-104	13.4 ± 0.26	a (-4)	37.9 ± 3.29	a (-11)	30.0 ± 0.61	ab (-4)
2018-2019		F-120	14.5 ± 0.45	a (+4)	48.3 ± 3.40	a (+13)	33.3 ± 1.13	a (+6)
		C-M	14.3 ± 0.61	ab	44.9 ± 5.28	ab	32.5 ± 1.56	ab
	LIDEA	F-96	13.2 ± 0.44	b (-8)	36.7 ± 4.39	b (-18)	30.0 ± 1.10	b (-8)
	UKLA	F-104	14.7 ± 0.32	a (+3)	49.2 ± 1.50	a (+10)	33.4 ± 0.61	a (+3)
		F-120	14.5 ± 0.19	ab (+1)	48.6 ± 0.80	a (+8)	32.8 ± 0.54	ab (+1)
		0N	9.9 ± 0.41		21.6 ± 2.66		15.8 ± 0.96	
		C-M	13.2 ± 0.33	b	32.9 ± 2.66	bc	26.8 ± 1.19	b
2019-2020	LIDEA	F-96	12.0 ± 0.15	c (-9)	29.1 ± 1.02	c (-12)	23.3 ± 0.48	c (-13)
	UKEA	F-104	13.9 ± 0.35	ab (+6)	40.4 ± 5.34	ab (+23)	30.0 ± 1.27	a (+12)
		F-120	14.2 ± 0.13	a (+8)	46.3 ± 0.69	a (+41)	31.0 ± 0.23	a (+16)

USING HUMIC COMPOUNDS TO IMPROVE EFFICIENCY OF FERTILISER NITROGEN



* https://www.massey.ac.nz/~flrc/workshops/13/Manuscripts/Paper_Schofield_2013.pdf

Figure 2. Pasture production at nine harvest dates from plots treated with dissolved urea and humic compounds or granular urea





* https://www.massey.ac.nz/~flrc/workshops/13/Manuscripts/Paper_Schofield_2013.pd

 Thank you, Questions?

 Mailing List, Resources, More info:

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Key Functions of Essential Nutrients

- N Chlorophyll, AA, P
- P Energy, root development
- K Enzyme production, sugar movement, N utilisation
- Ca Cell wall strength
- Mg Chlorophyll
- S N utilisation, root development
- Si cell wall strength

- B sugar translocation, reproductive processes
- Cu disease protection
- Zn auxin production, leaf size
- *Mn reproductive processes*
- Fe chlorophyll production
- Mo N utilisation
- Co N fixation
- Ni urease enzyme



Magnesium

- Central to chlorophyll (15-20%).
- Majority of Mg in the plant is used to catalyse protein synthesis (75%).
- Phosphorus synergist.





Manganese

- Splits water for photosynthesis (along with calcium).
- Important for seed health and germination.
- Key disease fighting nutrient.











Nitrogen Metabolism



Mineral		Current level			
Total Sugars	%	0.8			
	%	0.2 2			i
рн		5,8			
		5,5 "	·		
FC .	m\$/cm	12.2 1			1
	m\$/cm	17.7			
		11,1	/		
K - Potassium	ppm	3997 1			1
	ppm	4461 2			1
fa falster					
ca - calcium	ppm	551	_		
	ppm	1773 1		•	
K/Ca		7.25			1
		2 52 2	i		
		8,24			
Mg - Magnesium	ppm	380 '			1
	ppm	553 *			1
Na - sodium	ppm	11			
	ppm	35 '			
NH4 - Ammonium	nom	54 1		-	1
	ppm	69 8			
		~			
NO3 - Nitrate	ppm	3138			
	ppm	8032 2			
and the automotion					
N IS NICUSE	ppm	708			
	ppm	1813 '	1		
N - Total Nitrogen	000	1458			1
	0000	2222 8			
		1313			
CI - Chloride	ppm	417			1
	ppm	178 2	_		
S. Sulfur	2000	638 1			
5 5410	pperio	0.10			
	ppm	118/ 1	/	-	
P - Phosphorus	ppm	591 1			
	ppm	626 2			
5i - 5ilica	ppm	7,2			
	ppm	13.6 2)		
Fe - Iron	000	0.60			
	800	0.45 1			
		6,45			
Mn - Manganese	ppm	4,03			1
	ppm	7,86			1
To Tes		4.70			
211-2316	ppm	1,79			
	ppm	0.88 '			
8 - Boron	ppm	0.72 1	<u> </u>		1
	ppm	1.85			i
		4145			
Cu - Copper	ppm	1,84			
	ppm	3,20 2			
Mo - Mohdonum	2000	0.01			1
Mo · Moryodenum	ppm	0,06			
	yşım	0,05			
Al - Aluminium	ppm	<0.50 ¹	1		
	ppm	(0.50	i		i











Metabolic Shortcutting



Plant, Cell & Environment	PC GE
Original Article 🙃 Open Access	
The carbon bonus of organic nitrogen enhances nitrogen use efficiency of plants	

Oskar Franklin 🕿, Camila Aguetoni Cambui, Linda Gruffman, Sari Palmroth, Ram Oren, Torgny Näsholm

First published: 31 May 2016

The importance of organic nitrogen (N) for plant nutrition and productivity is increasingly being recognized. Here we show that it is not only the availability in the soil that matters, but also the effects on plant growth. The chemical form of N taken up, whether inorganic (such as nitrate) or organic (such as amino acids), may significantly influence plant shoot and root growth, and nitrogen use efficiency (NUE). We analysed these effects by synthesizing results from multiple laboratory experiments on small seedlings (Arabidopsis, poplar, pine and spruce) based on a tractable plant growth model. A key point is that the carbon cost of assimilating organic N into proteins is lower than that of inorganic N, mainly because of its carbon content. This carbon bonus makes it more beneficial for plants to take up organic than inorganic N, even when its availability to the roots is much lower – up to 70% lower for Arabidopsis seedlings. At equal growth rate, root-tratio was up to three times higher and nitrogen productivity up to 20% higher for organic than inorganic N, which both are factors that may contribute to higher NUE in crop production.

* Franklin, O., et al., (2016). doi: 10.1111/pce.12772



Table 2. Biochemically calculated assimilation costs for different N sources in gC gN^{-1} according to Zerihun *et al.* (1998)

N source	Gross C costs	C bonus ^a	Net N assimilation C cost
NO ₃	5.81	0	5.81
NH ₄	4.32	0	4.32
Gln	4.30	2.14	2.16
Arg	4.30	1.29	3.02

* Franklin, O., et al., (2016). doi: 10.1111/pce.12772

^a C bonus is equal to the molecular gC per gN.
^b Calculated assuming gross C costs (without C bonus) for N assimilation are equal to GIn.



Molybdenum

- Protein synthesis
- 2 key enzymes convert nitrate into nitrite then into ammonia/ammonium before it is used to synthesise amino acids.
- N_2 fixing bacteria also require Mo (& Fe) to access N_2 gas from the air.





Nickel

- Ni is part of the *urease enzyme* which splits the urea molecule liberating the N for plant metabolism.
- Without Ni/urease, urea can build up in plant tissues and become toxic.
- Plants specifically fed urea without Ni can be 'functionally N deficient'
- *Nickel sulphate* at 0.2% solution in barley around 20-50 g/ha can be effective.



Sulphur

- Amino acids
- Protein synthesis
- Root development
- Disease resistance
- Nodulation in legumes





Zinc



- Zinc determines **leaf size** solar panel.
- Chlorophyll synthesis.
- Nitrogen metabolism.



Copper

- Cu responsible for **lignin** production (primary defence) and **antimicrobial** compounds (secondary defence).
- Disease resistance.
- Metabolism of proteins and carbohydrates.
- Respiration.



Calcium

- Ca is deposited in **cell walls** and improves nutrient uptake of all elements into the cells.
- Ca is a cell strengthener (along with B & Si) improving pest and disease resistance.
- Highly immobile (deficiency symptoms on young leaves).



Boron

- Much plant B is also found deposited in cell walls.
- Plays a key role in synthesis of structural compounds (lignin, polyphenols) = primary defences.
- Root growth.
- Growing tips (very immobile).
- Reproductive processes!



Cobalt

- Vitamin B12 cell division.
- N fixing in legumes.
- Inputs
 - Cobalt sulphate





- Speed of absorption aa > urea > ammonium > nitrate.
- Protein hydrolysates and urea are ideal.
- Urea can improve uptake of trace elements when combined together Zn.

In Summary – Principles

- Plants can make use of many N forms manage them all.
- Different N forms work synergistically combine iN and oN.
- C-based N (oN) is more efficient for plant metabolism.
- N is not an island needs other synergistic nutrients.
- Excess N (or imbalanced N) compromises plant health.
- Foliar N (aa & urea) rapidly absorbed, less losses, lower rates, efficiency gains.

In Summary – Practice

- Nitrate best used during vegetative stages only.
- High analysis, embedded C and rapid absorption gives urea a key advantage for foliar applications.
- Include nutrient synergists ideally determine via plant analysis.
- Include carbon and pH modification.

Witte, C.P. (2011). doi: 10.1016/j.plantsci.2010.11.010







Foliar N Combinations

- General/All purpose:
 - Urea at ~20 kg/ha
 - Multi-trace element package at label rates

- Carbon source, pH adjustment, wetter/sticker*



Foliar N Combinations

• Photosynthesis Primer:

- Urea at ~20 kg/ha
- Magnesium sulphate at 5-7 kg/ha
- Iron sulphate at 0.5-1 kg/ha
- Manganese sulphate at 0.5-1-2 kg/ha
- Carbon source, pH adjustment, wetter/sticker*

Foliar N Combinations

• Grain Fill:

- Urea at ~20 kg/ha
- Potassium sulphate at 3-5 kg/ha
- Carbon source, pH adjustment, wetter/sticker*

Organic Foliar N

- Certified Organic
 - Protein hydrolysate at 3-10 L/ha
 - Approved amino acid product at label rates or protein hydrolysate at ~5 L/ha
 - Trace elements (use plant analysis to confirm limitation)

- Carbon source, pH adjustment, wetter/sticker*

Trace Elements

Zinc sulphate [Zn]1 kg/ha (lbs/ac)Iron sulphate [Fe]1 kg/ha (lbs/ac)Manganese sulphate [Mn]1 kg/ha (lbs/ac)Copper sulphate [Cu]0.5-1 kg/ha (lbs/ac)Sodium borate [B]0.5-1 kg/ha (lbs/ac)Sodium molybdate [Mo]50-200 g/ha (1.7-7 oz/ac)Nickel sulphate [Ni]50-100 g/ha (1.7-3.5 oz/ac)Cobalt sulphate [Co]100 g/ha (3.5 oz/ac)

Cropland: 1% solution [1 lbs per 12 gal of water]

• Horticulture: 0.1% solution [3.5 oz per 12 gal of water]

Timings – Cereals

- Just ahead of flag leaf
- End of flowering
- (tillering, stem elongation and booting also somewhat beneficial)



Timings – Canola

- Pre flowering
- Post flowering
- Include sulphur –Urea + SOA etc



Timings – Pulses

- After flowering during pod set
- 5-10 kg/ha



Grasslands

- 20-25 kg/ha (lbs/ac)
- Dairy

 Every 3-4 weeks
 Around 7 days after each grazing event
- Beef -3 to 4 applications per growing season



Solubilising Urea Tips

- Melting urea is an endothermic reaction, warm water helps.
- Leave water in tank for a few days to allow sun warming black tank etc.
- Slowly feed the urea with an auger/conveyor into moving water, the more circulation the better.
- Direct the return of the circulation down and around the bottom of the tank preventing any urea to settle (can become difficult to solubilise).

Minimising Scorch

- If dissolving urea, be sure it is low biuret (<1%).
- Nickel sulphate at ~20-50 g/ha
- Apply with high humidity (evening, night, heavy dew).
- Avoid bright sunny days and windy conditions.
- Low spray pressure (<50 psi)
- Low temperatures (<0C) or significant temperature fluctuations can increase burn.
- Dilute with additional water if necessary.





* Paynel, F., et al (2008). doi: 10.1051/agro:2007061

Ecosystems Environme

Grass-legume mixtures can yield more nitrogen than legume pure stands due to mutual stimulation of nitrogen uptake from symbiotic and non-symbiotic sources

Concerted use of legumes and of functional diversity in grassland forage systems can provide major contributions to the challenges of agricultural systems being productive yet environmental friendly. Acquisition and transformation of nitrogen (N) resources by legumes and grasses were studied in a temperate grassland experiment near Zurich (Switzerland) to investigate mechanisms driving effects of functional diversity in mixed swards and to optimise mixtures for efficient resource use.

Grass-legume interactions and N availability were varied by manipulating legume percentage of the sward (0-1003) and N fertiliser application (50, 150 or 450 kg of N ha⁻¹ year⁻¹).¹⁵N technology quantified N derived from symbiotic (Nsym) and non-symbiotic (Nnonsym) sources. Generally, acquisition of Nsym by the entire mixture was stimulated by grasses, As a result, strong

Cenerally, acquisition of Nsym by the entire mixture was stimulated by grasses. As a result, strong overyielding of Nsym occurred (e.g. 75 and 114% for year 1 and 2 at 1150) and mixtures with only 60% and 37% legumes (year 1 and 2) already attained the same Nsym yield as pure legume stands. Legumes stimlated Nonsym acquisition by the entire mixture, largely via increased uptake by the grass component. Thus, overyielding of Nonsym of 31% occurred in year 1 (N150). Mutual grass-legume interactions stimulated acquisition of Nsym, acquisition of Nonsym and effi-

Mutual grass-legume interactions stimulated acquisition of Nsym, acquisition of Nnonsym and efficient transformation of N into biomass compared to either monocultures. These effects of functional diversity can substantially contribute to productive and resource efficient agricultural grassland systems and were maximised in mixtures with 40–602 (gumes.

We describe how legume proportion modifies N acquisition from different sources.

Symbiotic N₂ fixation was stimulated in mixtures compared to monocultures.

Uptake of N from soil N pools was stimulated in mixtures compared to monocultures.

The acquired N was used more efficiently by mixtures for biomass production.



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Yield of binary- and multi-species swards relative to single-species swards in intensive silage systems



* Moloney, T., et al., (2020). doi: 10.2478/ijafr-2020-000

Defining NUE

- Nitrogen uptake efficiency (NUpE) is defined as the total N in plants relative to the applied N fertilizer.
- Nitrogen utilisation efficiency (NUtE) is defined as plant biomass or seed yield relative to total N in the plant, reflecting the capacity of plants to convert acquired N to plant biomass or seed yield.

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Yield of binary- and multi-species swards relative to single-species swards in intensive silage systems



* Moloney, T., et al., (2020). doi: 10.2478/ijafr-2020-0002

siology and Fertility of **Soils**

Catch crop diversity increases rhizosphere carbon input and soil microbial biomass

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- Mustard (mono) vs 4-way mix or 12-way mix
- CO₂ uptake 2x higher in mix-4 and 3x higher in mix-12
- Total microbial biomass 8% higher in mix-4 and 18% higher in mix-12
- Fungal and actinobacteria most responsive
- C residence time increased with mixture by up to 1.8 times



The results of this study suggest positive impacts of plant diversity on C cycling by higher atmospheric C uptake, higher transport rates towards the rhizosphere, higher microbial incorporation and prolonged residence time in the soil environment.





